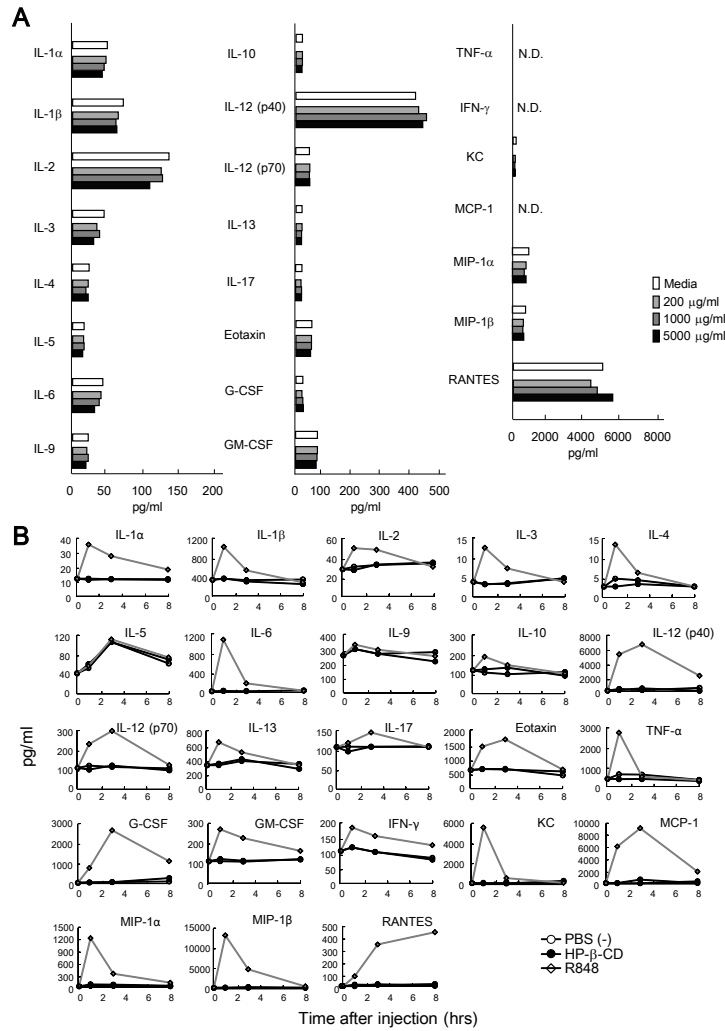
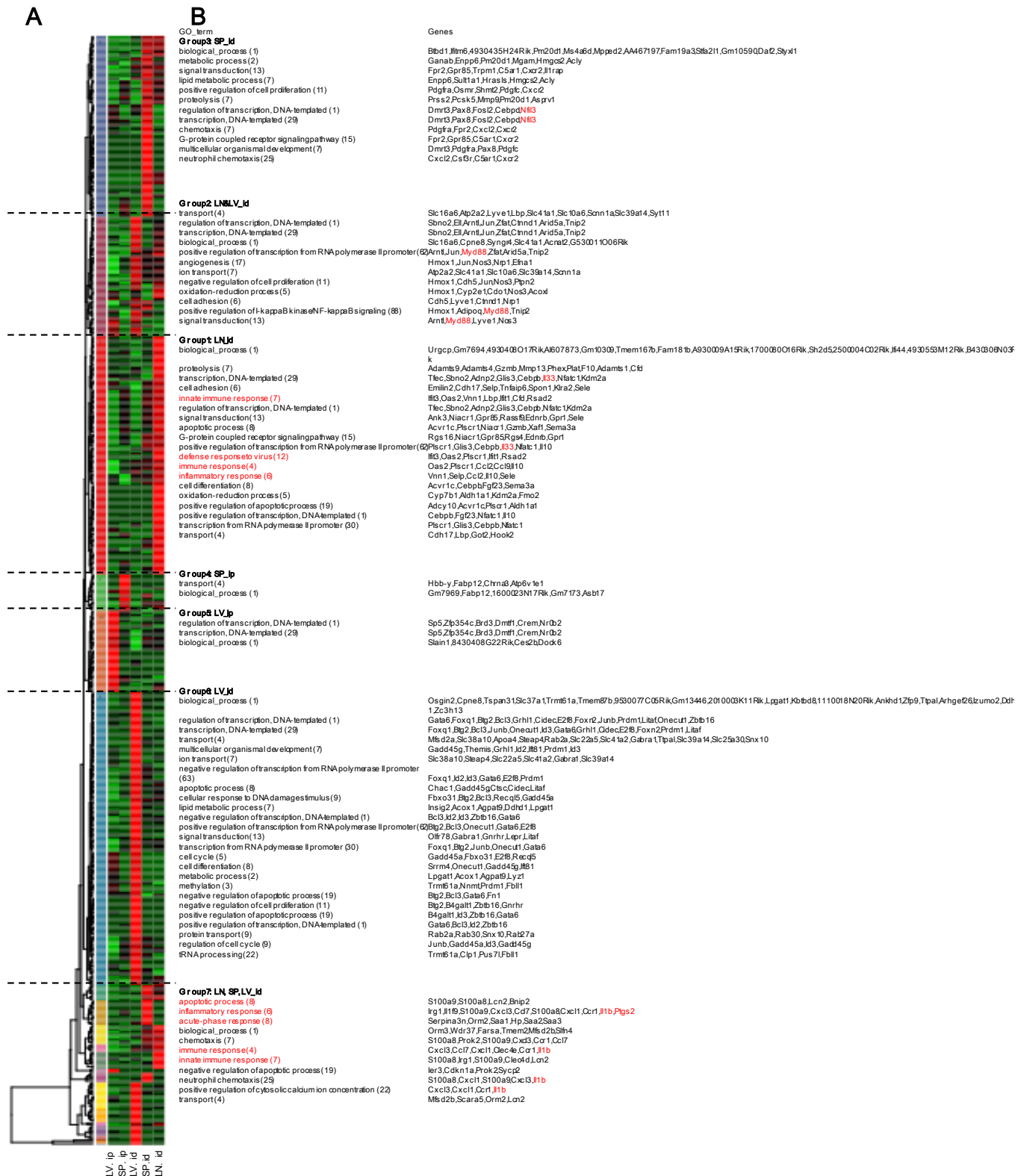


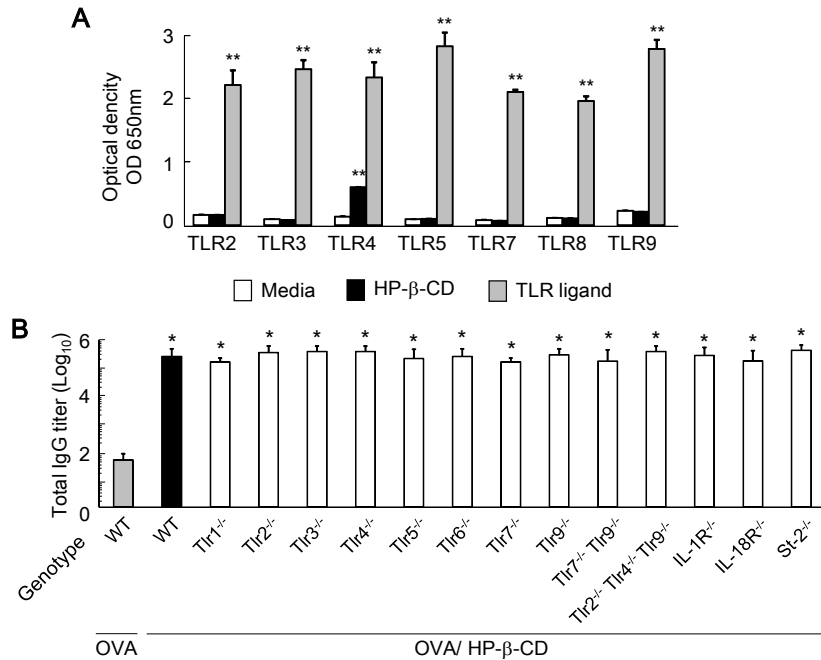
Supplemental figures.



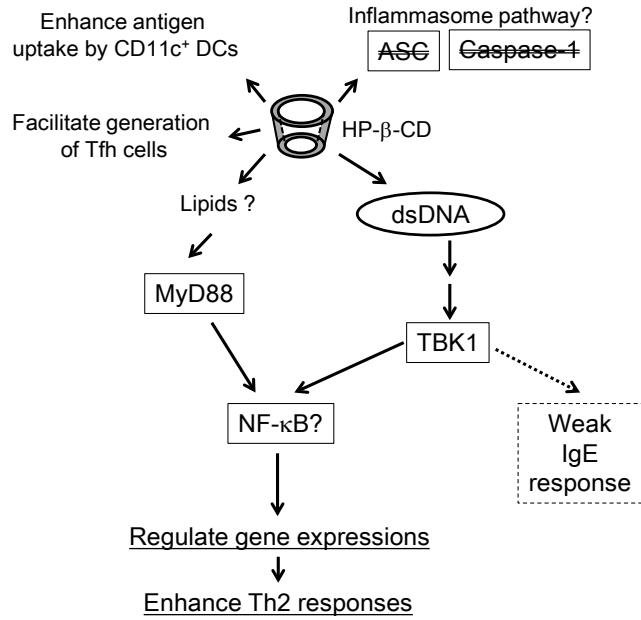
**Supplemental Fig. 1. The potential of HP-β-CD to induce proinflammatory cytokine responses *in vitro* and *in vivo*.** (A) Evaluation of the cytokine responses in splenocytes stimulated with HP-β-CD. Splenocytes were stimulated with HP-β-CD and the supernatant was collected 24 h after stimulation. The concentration of each cytokine in the supernatant was measured with the Bio-Plex Pro™ Mouse Cytokine 23-plex Assay (Bio-Rad Laboratories, Inc.). Data are the averages of two independent experiments. (B) Evaluation of the cytokine responses in mice injected with HP-β-CD. Mice were s.c. injected with 30% HP-β-CD or 100 μg of R848 (n = 3). The sera were collected 1, 3, and 8 h after injection, and those from three mice treated identically were pooled. Each cytokine concentration was measured with the Bio-Plex Pro™ Mouse Cytokine 23-plex Assay.



**Supplemental Fig. 2. Group clustering using mRNA expression levels and GO analysis of HP-β-CD-induced genes in different organs after its i.d. or i.p. administration.** (A) The expression levels were normalized using the Z-scores for the fold-change values. Upregulated genes were selected, under the condition of a fold change > 2 and a customized PA call = 1 in at least one sample, and clustered into seven groups using complete-linkage hierarchical clustering. (B) For each group, the clustered genes were annotated by their functional roles using the AMIGO gene ontology database, and representative GO terms are indicated. The number in parentheses next to the GO term in the column indicates its rank in the GO hierarchy, and relates to the number of ancestors of the given term (i.e., biological process (1) indicates the root of the GO hierarchy tree). The gene list in the column shows the GO-categorized genes found in each grouped cluster.



**Supplemental Fig. 3. Involvement of TLRs and IL-1R superfamily in the adjuvanticity of HP-β-CD** (A) TLR ligand screening with HEK-Blue™ TLR cells. HEK-Blue TLR cells (Invivogen) are NF-κB-secreting alkaline phosphatase reporter cells that stably express mouse TLR genes (*Tlr2*, 3, 4, 5, 7, 8, or 9). TLR stimulation was tested by assessing NF-κB activation in HEK293 cells expressing a given TLR. After incubation for 16–20 h with the test substance, the OD was measured at 650 nm on a Molecular Devices Spectra Max 340PC Absorbance Detector. HP-β-CD was tested at 5000 μg/ml. The following substances were used as positive controls: for TLR2: heat-killed *Listeria monocytogenes* (HKLM) at 10<sup>8</sup> cell/ml; for TLR3: poly(I:C) at 1 μg/ml; for TLR4: *E. coli* K12 LPS at 100 ng/ml; for TLR5: *Salmonella enterica* subspecies I, serovar *typhimurium* flagellin at 100 ng/ml; for TLR7: CL097 at 1 μg/ml; for TLR8: CL075 at 10 μg/ml + poly(dT) at 10 μM; and for TLR9: CpG ODN 1826 at 1 μg/ml. An assay was performed according to the manufacturer's instructions. Data are representative of two independent experiments. \*\*P < 0.01 v.s. media on Student's *t* test. (B) Evaluation of the humoral responses in KO mice lacking TLRs or IL-1R superfamily genes. Mice were s.c. immunized twice with OVA/30% HP-β-CD (n = 4–6). The total anti-OVA antibody titers were measured with an ELISA 7 days after the second immunization. Error bars denote SD. \*P < 0.05 v.s. WT mice immunized with OVA alone on Student's *t* test.



**Supplemental Fig. 4. Biological activities of locally injected HP-β-CD and the proposed immunological mechanisms/signaling pathways involved in the adjuvant activity of HP-β-CD.** 1) Antigen uptake: HP-β-CD enhances antigen uptake by CD11c<sup>+</sup> DCs; 2) Facilitates the generation of Tfh cells; 3) MyD88-dependent signaling pathway; 4) TBK1-dependent signaling pathway: the temporary release of dsDNA at the injection site activates the TBK1-dependent signaling pathway.

## Legend for Movie

**Movie S1. 3D reconstruction of the two-photon images of the LN. HP- $\beta$ -CD (green), OVA (blue), MARCO<sup>+</sup> macrophages (red) . Mice were i.d. injected with OVA–Alexa Fluor 647/ 3% HP- $\beta$ -CD -FITC, and after 30 min, anti-MARCO antibody was i.d. injected. LNs were collected 30 min after the antibody injection (i.d.), and were imaged by two-photon microscopy.**