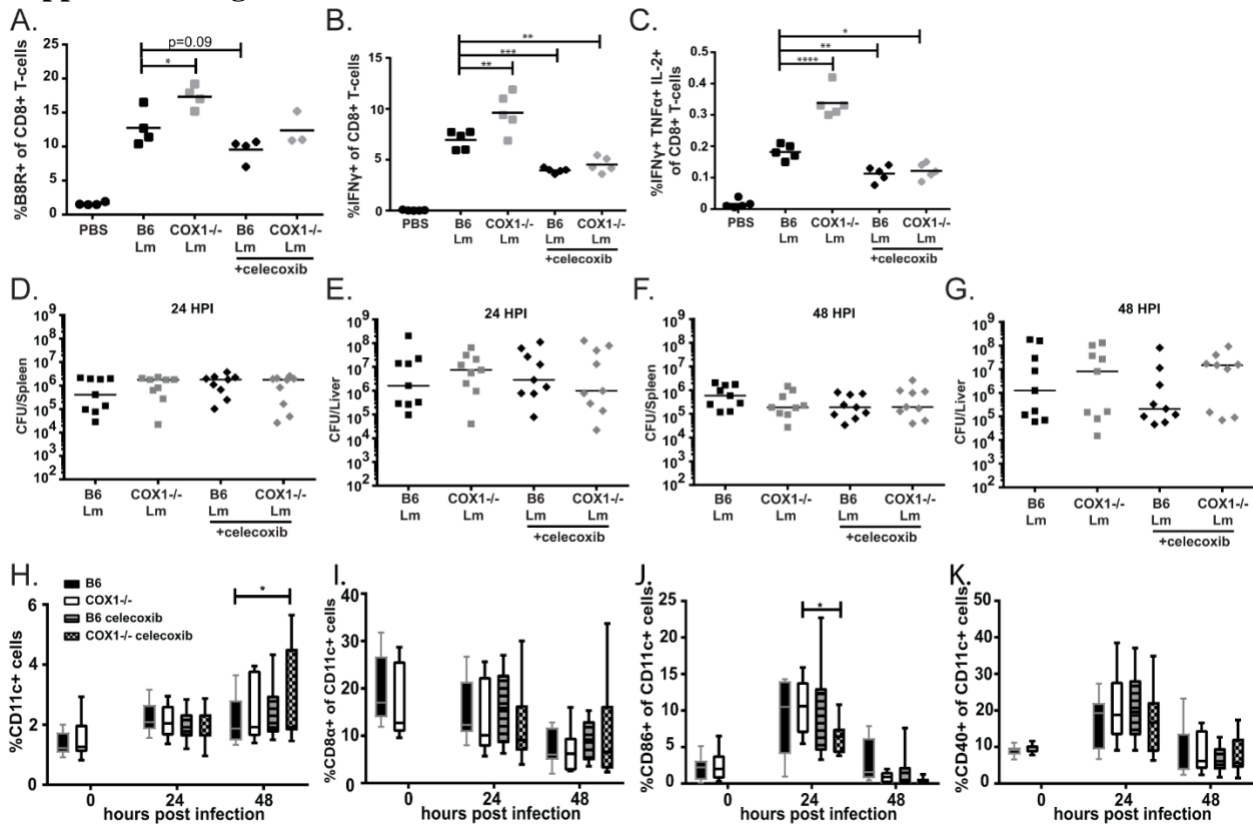


1 Supplemental Figures



2 Supplemental Figure 1: Cyclooxygenase modulation does not influence bacterial burdens or

3 DC maturation states. The indicated strains of mice in the presence or absence of celecoxib-laden

4 chow were immunized with 1×10^7 cfu attenuated Lm. CD8+ T-cells were assessed 7 days post

5 immunization by B8R tetramer (A), or by OVA-stimulated IFN γ (B) or IFN γ , TNF α , and IL-2

6 (C). Bacterial burdens in the spleens or livers were assessed 24 or 48 hours post immunization.

7 Spleens and livers were removed, homogenized and plated for bacterial burdens (D-G). Spleens

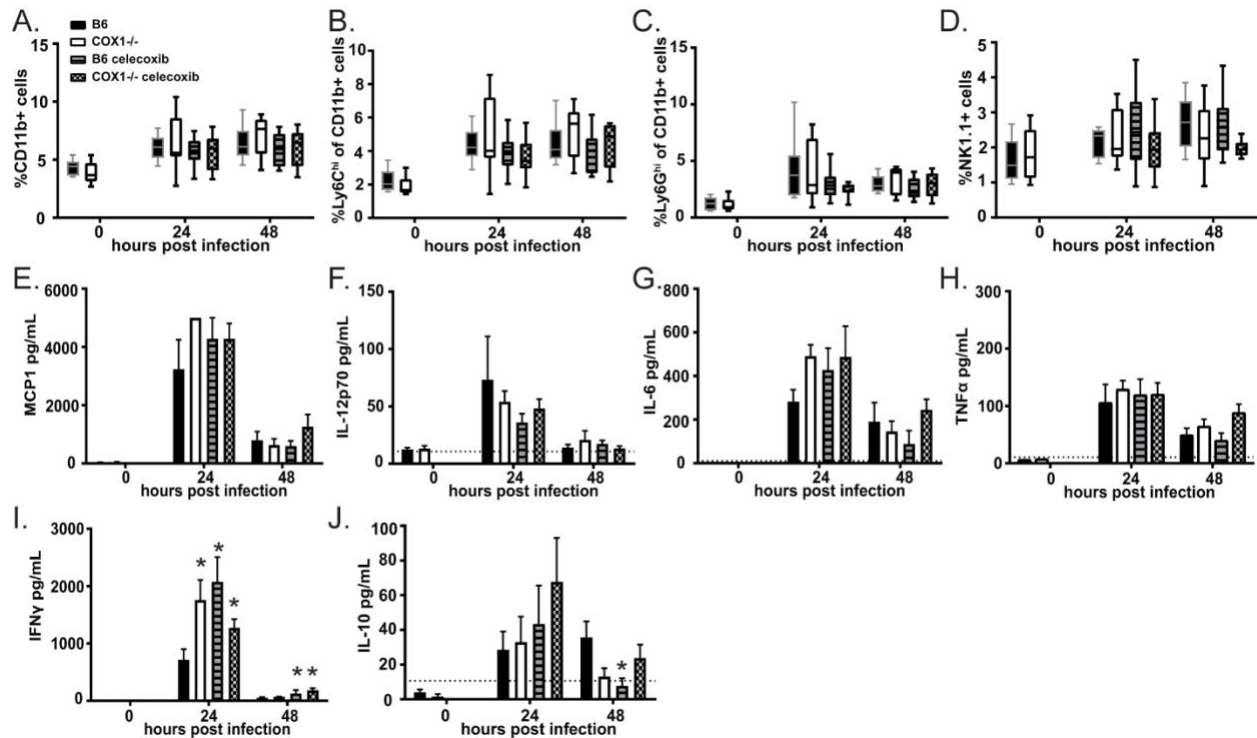
8 were removed and analyzed by flow cytometry (H-K). Samples were gated on CD11c+ (H-K),

9 followed by selection for CD8 α (I), CD86 (J) or CD40 (K). Data are representative of 2

10 independent experiments of 5 mice per group (A-C) or are the combination of 3 independent

11 experiments of 3 mice per group. Significance was determined by a one-way ANOVA with

12 Bonferroni's correction. * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$



13

14 Supplemental Figure 2: Cyclooxygenase modulation does not impact innate immune cell

15 **recruitment, but influences innate cytokine production.** The indicated strains of mice in the

16 presence or absence of celecoxib-laden chow were immunized with 1×10^7 cfu attenuated Lm. 24

17 or 48 hours post immunization, spleens were removed and analyzed by flow cytometry (A-D).

18 Samples were gated on CD11b⁺ (A-C) followed by selection for Ly6C^{hi} populations (B) or Ly6G^{hi}

19 populations (C). Samples were gated for NK1.1 (D). 24 or 48 hours post immunization serum was

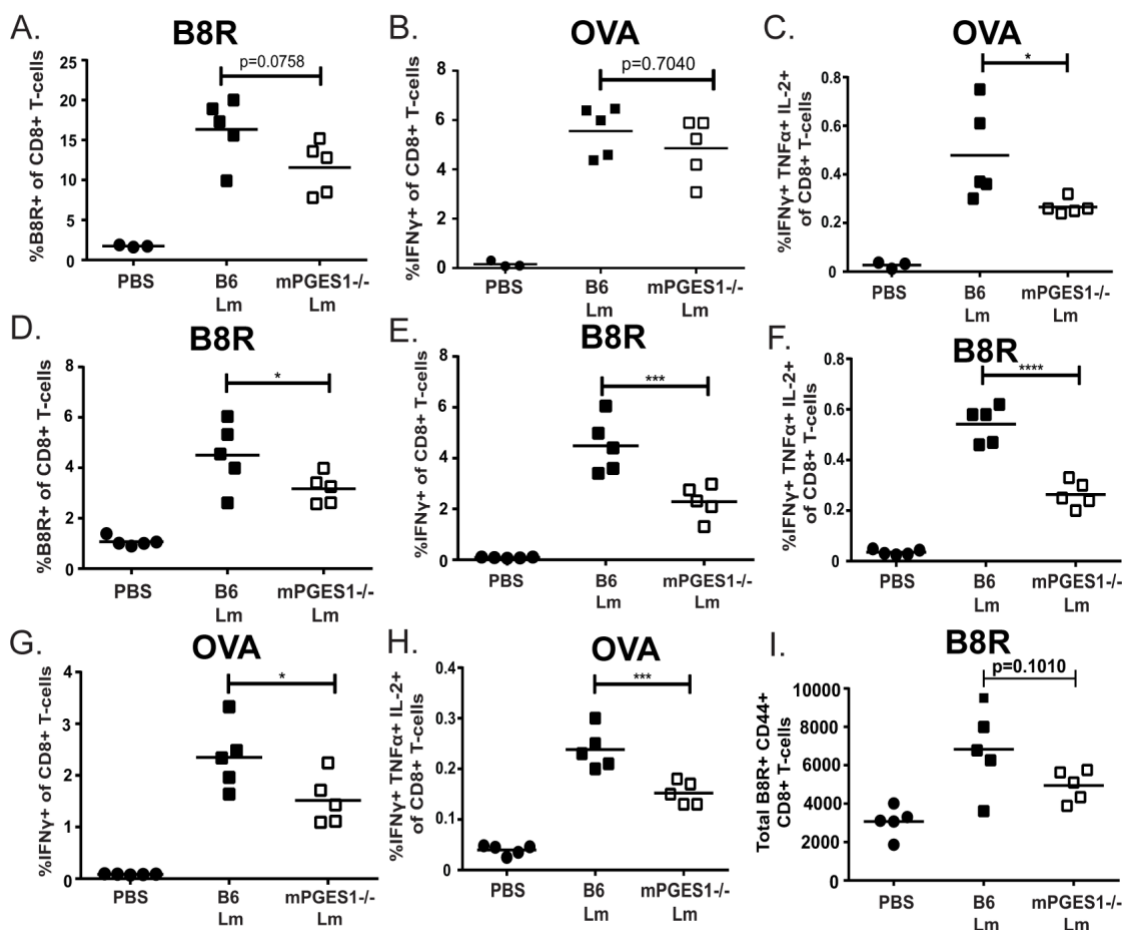
20 examined via cytokine bead array (E-J) for MCP-1 (E), IL-12p70 (F), IL-6 (G), TNF α (H), IFN γ

21 (I) or IL-10 (J). Data are of the combination of 3 independent experiments of 3 mice/group.

22 Significance was determined by a one-way ANOVA with Bonferroni's correction. * $p < 0.05$

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25

26 **Supplemental Figure 3: mPGES1^{-/-} mice have impaired cell-mediated immunity.** The

27 indicated strains of mice were immunized with either 1x10⁷ cfu (A-C) or 1x10³ cfu (D-H) of

28 attenuated Lm and assessed for the indicated antigen-specific response at 7 days post immunization

29 by tetramer (A, D) or by intracellular cytokine staining (B-C, E-H). Cells were gated for CD8+ T-

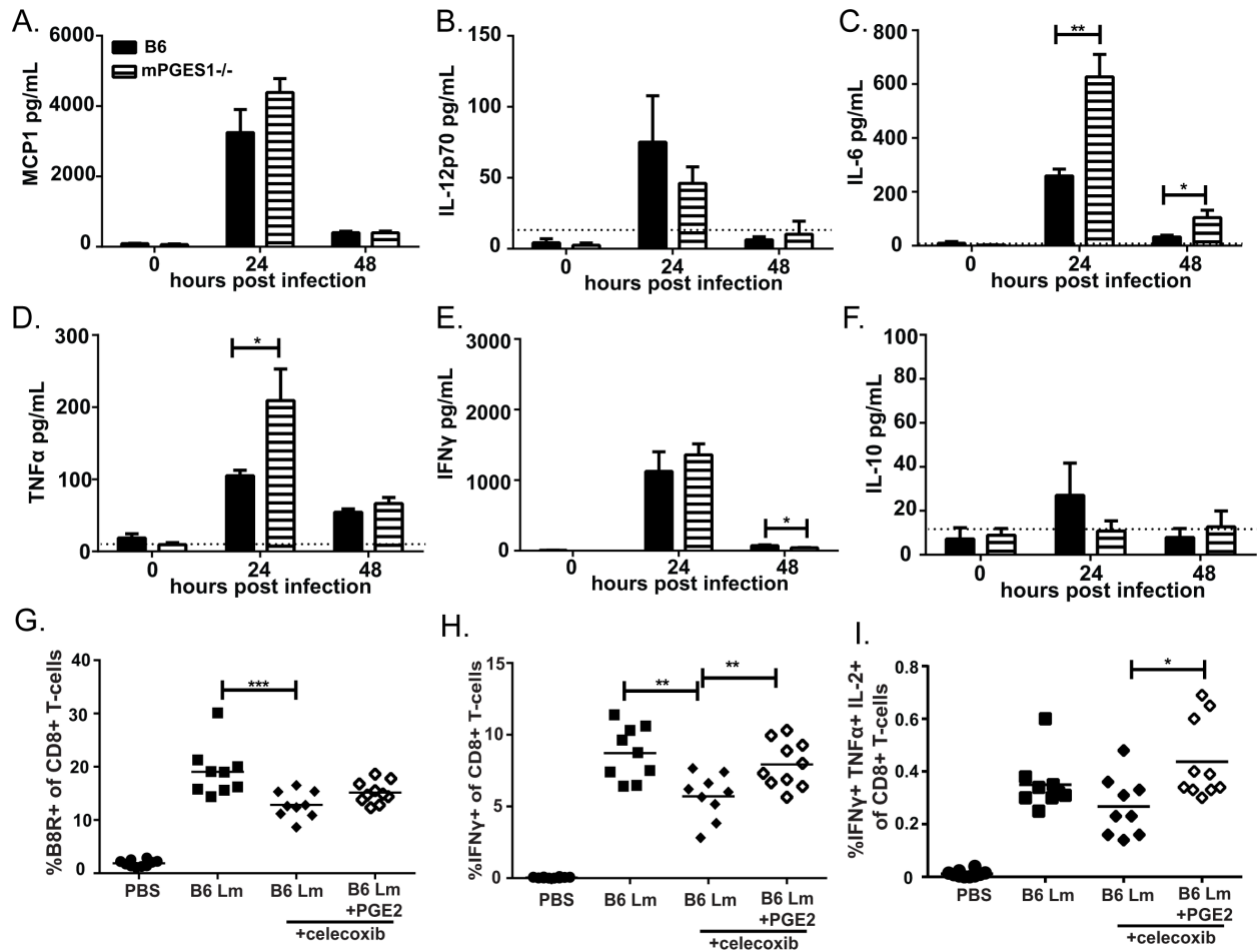
30 cells followed by IFN γ , TNF α , and IL-2 as indicated. Memory responses (I) were assessed

31 following immunization with 1x10⁷ cfu of attenuated Lm at 30 days post immunization via B8R-

32 tetramer staining on CD3+ CD8+ CD44+ cells. Data are representative of 2-3 independent

33 experiments of 5 mice per group. Significance was determined by a one-way ANOVA with

34 Bonferroni's correction. *p<0.05, **p<0.01, ***p<0.001



35

36 **Supplemental Figure 4: PGE2 is sufficient for immunity to *L. monocytogenes*.** Wild-type or
 37 mPGES1^{-/-} mice were immunized with 1×10^7 attenuated Lm. 24 or 48 hours post immunization
 38 serum was examined via cytokine bead array (A-F) for MCP-1 (A), IL-12p70 (B), IL-6 (C), TNFα
 39 (D), IFNγ (E) or IL-10 (F). PGE2 was supplemented in celecoxib treated animals immunized with
 40 1×10^7 cfu attenuated Lm (G-I) and primary responses were assessed via B8R-tetramer (G) or
 41 OVA-specific intracellular cytokine responses (H-I). Data are the combination of 2 independent
 42 experiments of 3-5 mice/group. Significance was determined by a student's t-test (A-F) or one-
 43 way ANOVA with Bonferroni's correction (G-I). *p<0.05, **p<0.01, ***p<0.001