

## Figure Legends for Supplemental Figures

Fig. S1. Blocking of targeting NY-ESO-1 protein binding to the cell surface of mo-DC by competitive antibody. (a) Binding of DEC-ESO and MR-ESO on mo-DC was investigated by flowcytometry. Mo-DC were preincubated with or without 10  $\mu\text{g/ml}$  DEC-hCG $\beta$  or MR-hCG $\beta$  on ice for 30 minutes and then incubated with 4  $\mu\text{g/ml}$  DEC-ESO (left) or MR-ESO (right). After extensive washes, NY-ESO-1 protein bound on mo-DC was detected by anti-NY-ESO-1 mAb and PE-conjugated anti-mouse IgG1 mAb. Similar results were obtained in mo-DCs from two donors. (b) Inhibition of cross-presentation of DEC-ESO and MR-ESO by blocking their binding to the cell surface molecules. Mo-DCs were preincubated with or without 100  $\mu\text{g/ml}$  DEC-hCG $\beta$  or MR-hCG $\beta$  on ice for 30 minutes and then incubated with 4  $\mu\text{g/ml}$  DEC-ESO or MR-ESO on ice for 15 minutes. After extensive washes, cells were incubated overnight at 37°C to allow the antigen processing and used as target cells in ELISPOT assay. Experiments were repeated twice and the result from one experiment was shown as mean + SD from duplicate samples. \*,  $p < 0.05$ .

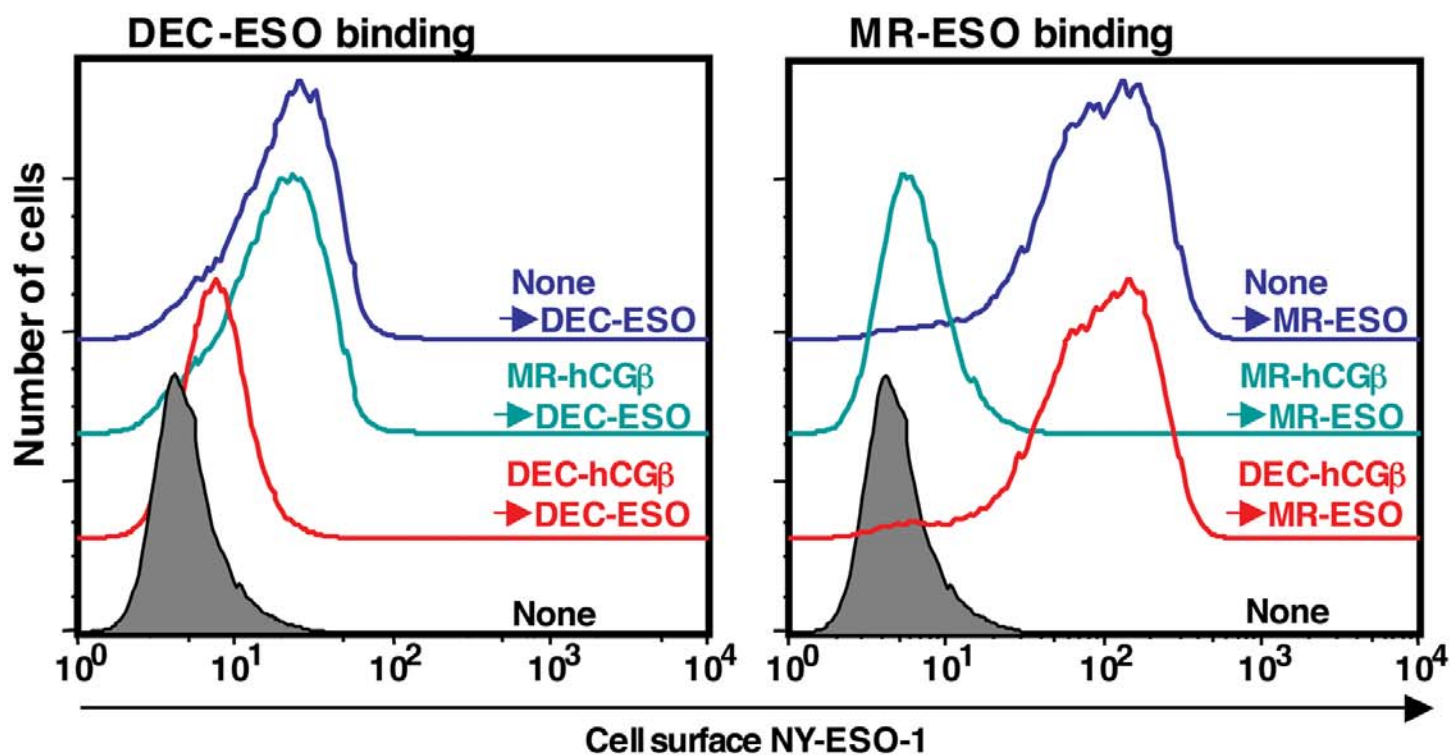
**Fig. S2.** Absence of stimulatory/costimulatory effect of targeted NY-ESO-1 proteins on monocyte-derived dendritic cells (mo-DCs). Mo-DCs were prepared as described in Materials and Methods. At day 6, mo-DCs were incubated with indicated targeted NY-ESO-1 fusion protein (DEC-ESO or MR-ESO) or recombinant NY-ESO-1 protein for 30 minutes before adding indicated toll like receptor ligands. After 24 hours, cells were harvested and stained with PE-conjugated anti-CD83 and FITC-conjugated anti-CD86 mAbs. Fluorescent intensity was measured and analyzed by a FACSCalibur instrument and CellQuest software, respectively.

**Fig. S3.** Absence of stimulatory/costimulatory effect of targeted NY-ESO-1 proteins on monocyte-derived dendritic cells (mo-DCs). Mo-DCs were prepared and stimulated as in Fig. S1. After 24 hours, supernatant was harvested and cytokine levels in the supernatant were measured by enzyme-linked immunosorbent assay (ELISA). The limit of detection in these assays was 78 pg/ml for IL-12, 9.4 pg/ml for IL-10, and 390 pg/ml for TNF. Experiments

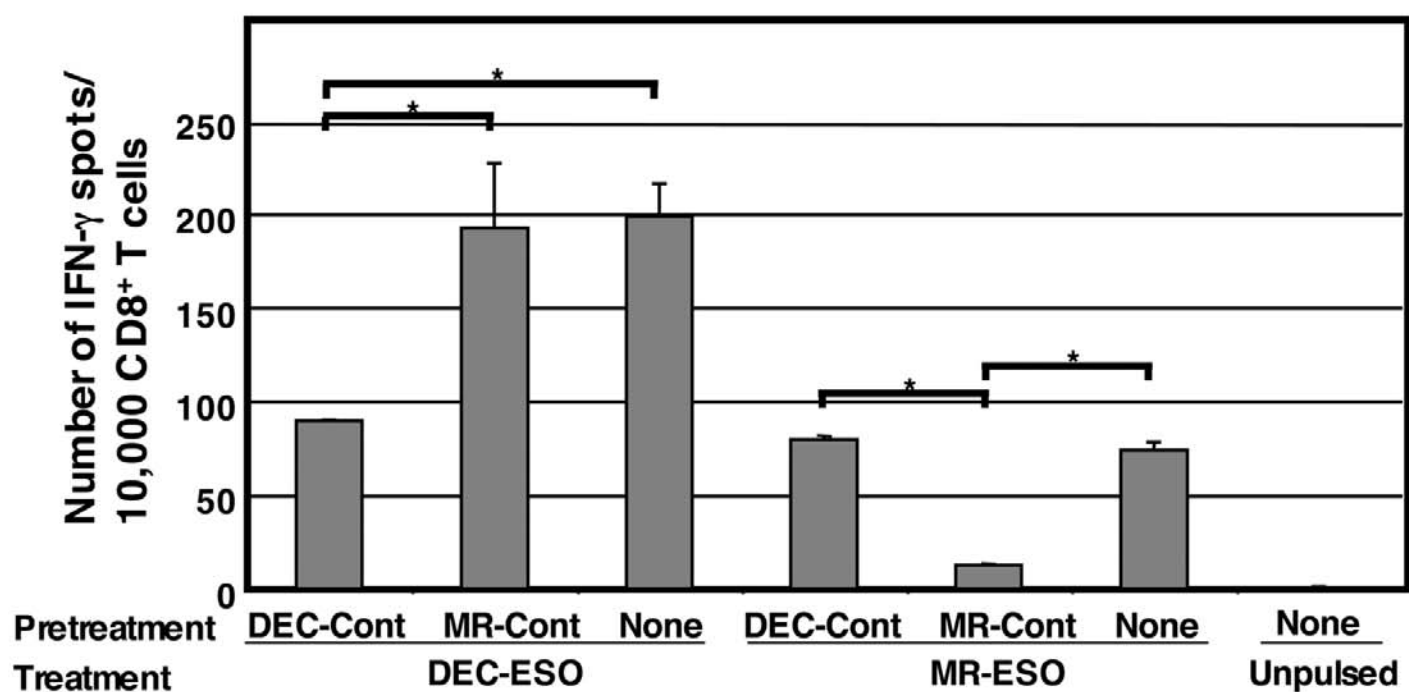
were performed using mo-DCs from two different healthy donors. The results were shown as mean + SD from duplicate samples.

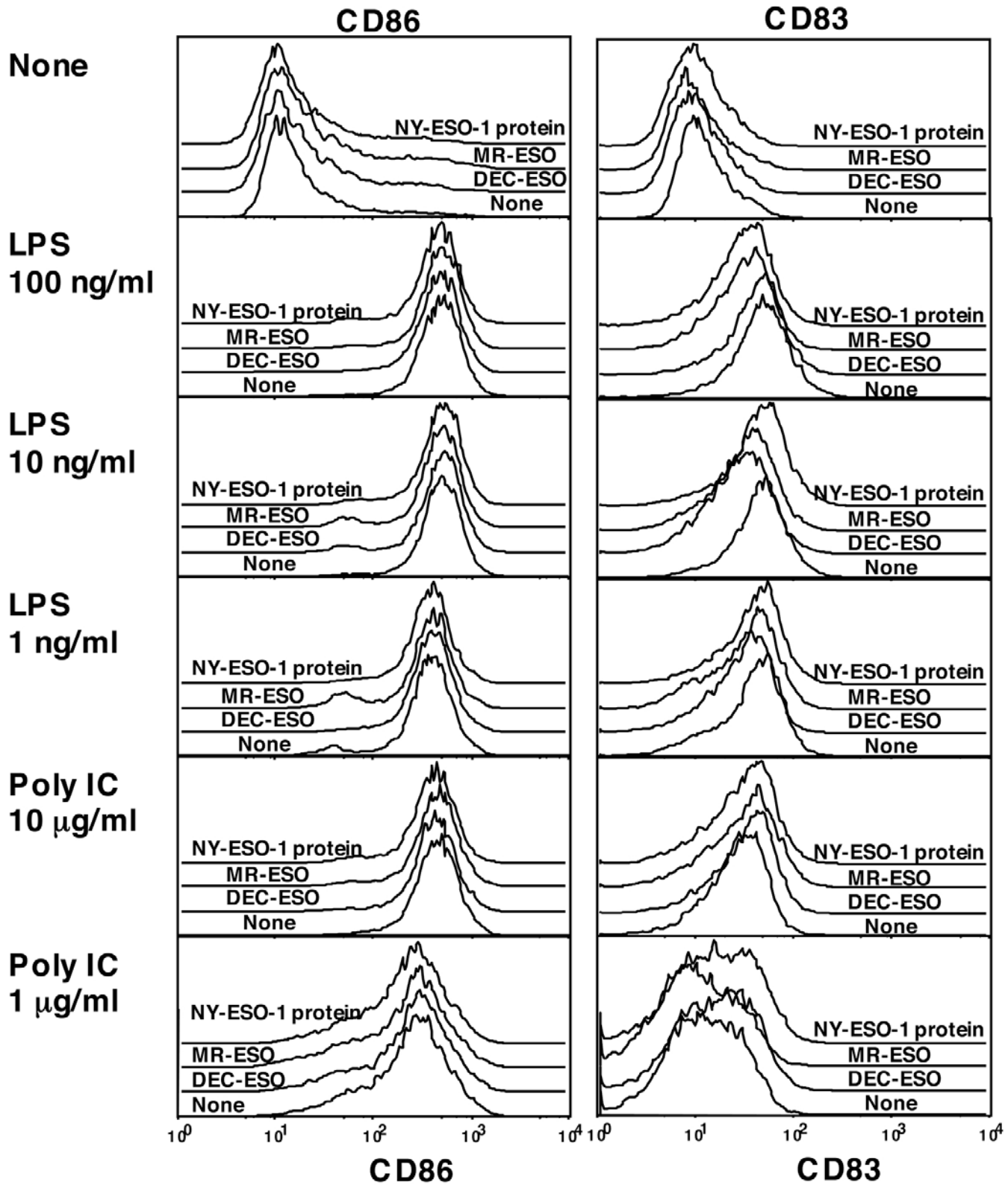
**Fig. S4.** Characterization of NY-ESO-1-specific CD8<sup>+</sup> T cell responses from patient AA. CD8<sup>+</sup> T cells from patient AA were presensitized and NY-ESO-1-specific CD8<sup>+</sup> T cell line was generated as described in Materials and Methods. (A-B) Peptide specificity of NY-ESO-1-specific CD8<sup>+</sup> T cell lines was evaluated using autologous EBV-B cells as APCs by ELISPOT assay. (C) Avidity of CD8<sup>+</sup> T cell line against short peptides was evaluated by ELISPOT assay using autologous EBV-B cells pulsed with an indicated concentration of short peptides. (D) HLA-restriction of NY-ESO-1<sub>101-109</sub> and NY-ESO-1<sub>102-110</sub> was determined as HLA-B\*4002 by HLA-compatible allogeneic EBV-B cells. (E) Tumor recognition of HLA-B\*4002-restricted and NY-ESO-1<sub>101-109</sub>-specific CD8<sup>+</sup> T cell line. SK-MEL-52: NY-ESO-1<sup>+</sup>HLA-B\*4002<sup>+</sup>; SK-MEL-139: NY-ESO-1<sup>+</sup>HLA-B4002<sup>-</sup>; SK-MEL-149: NY-ESO-1<sup>-</sup>HLA-B\*4002<sup>+</sup>; SK-MEL-37: NY-ESO-1<sup>+</sup>HLA-B4002<sup>-</sup>.

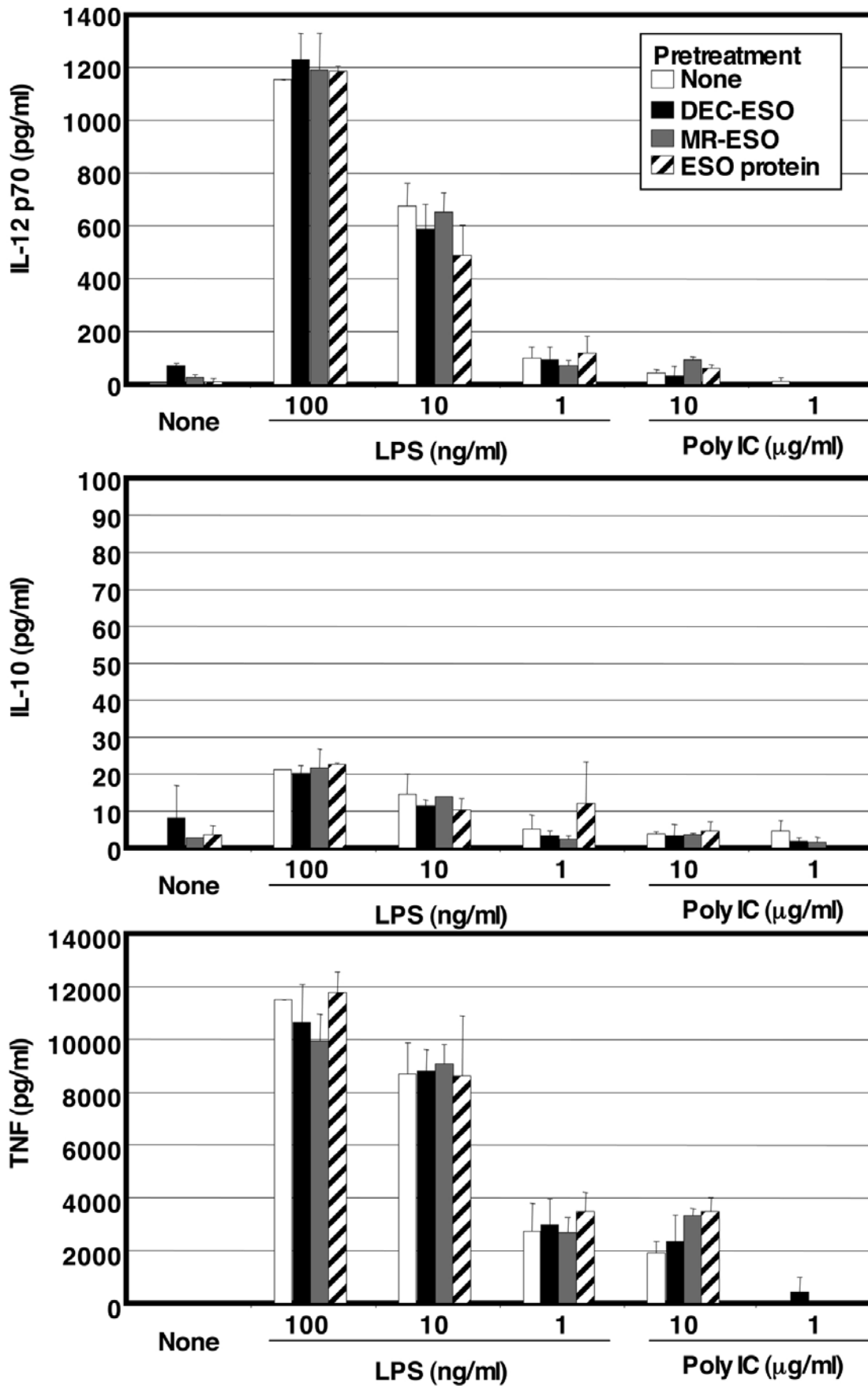
(a)



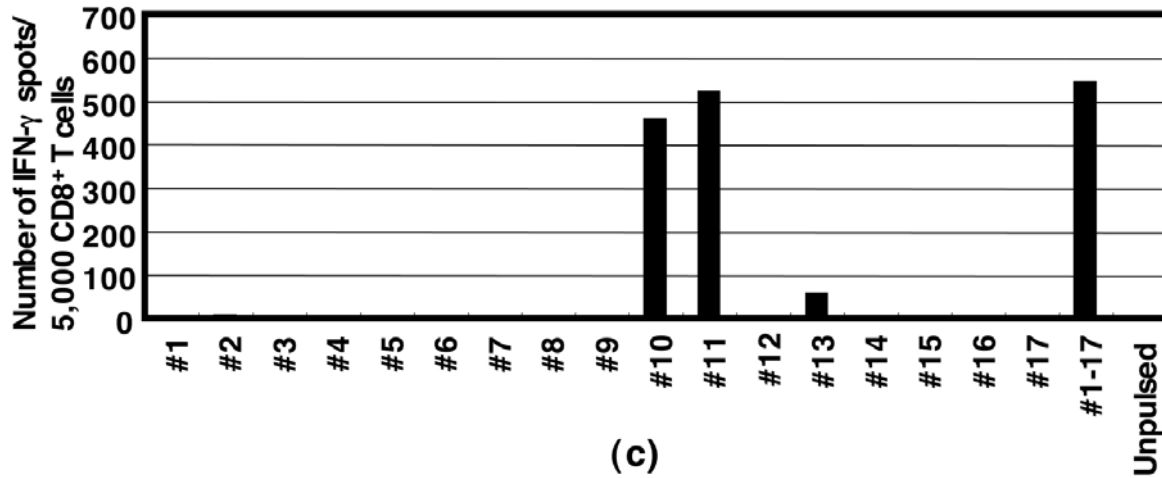
(b)



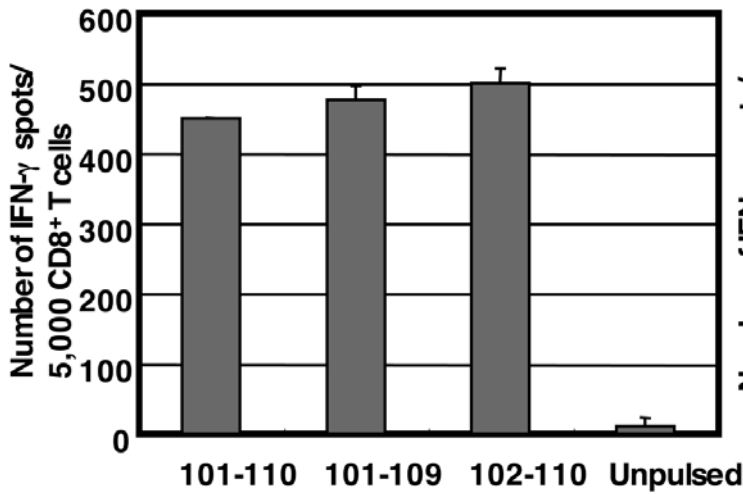




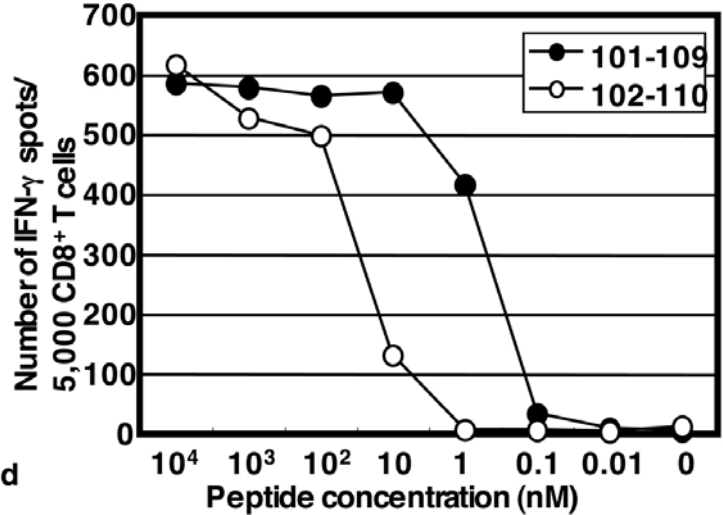
(a)



(b)

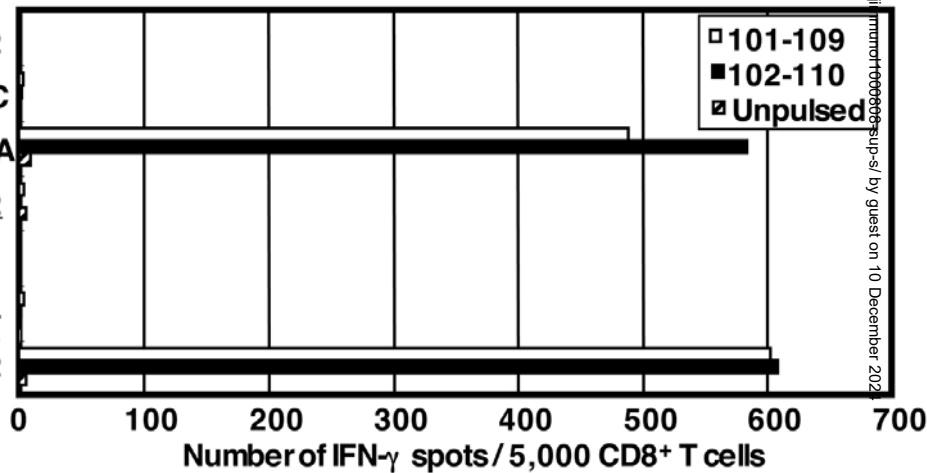


(c)



(d)

	HLA-A	HLA-B	HLA-C
NC116	2301 <u>2402</u>	42013906	17XX0702
NC214	26JV <u>3201</u>	40CCP 5001	030406BC
NC158	030131AB	3512 <u>4002</u>	030504APA
CB	020102XX	18015101	<u>02021202</u>
NC122	11012601	35033801	<u>1203</u>
NC136	<u>2402</u>	38011501	12NP0304
AA	24023201	40025101	12030202



(e)

